



Instructions for Catalog # 083A  
WasteWatR™ Coliform MicrobE™ - SM9221  
Revision 090119

Description:

- This standard consists of two glass vials each containing one gelatin tablet and a desiccant pouch (Sample A and B). The bacteria are contained in the gelatin tablets. Two sterile, 100 mL hydrating fluids are also provided. Sample A contains a certified number for both Colony Forming Units (CFU) and Most Probable Number (MPN) index and Sample B contains no microorganisms.
- This standard is not preserved.
- The glass vials containing the bacteria should be stored at  $4\pm 2^{\circ}\text{C}$ .
- The hydrating fluid can be stored at room temperature.
- This product is intended to be used as a quality control check of the entire analytical process for the analytes/matrix included in the standard for SM 9221 or equivalent multiple tube fermentation methods.
- ERA suggests that when subsampling this product prior to analysis you use a minimum sample size of at least 50 mL. Using a smaller sample size may invalidate the assigned value and/or uncertainty shown on the certificate of analysis.
- The certified values apply to the diluted sample after following the stated dilution instructions.

Helpful Hints:

- The bacteria are in a lyophilized form in the gelatin tablets and each sample must be hydrated per the following instructions prior to analysis.
- ERA recommends that the quality control guidelines in *Standard Methods for the Examination of Water and Wastewater*, Section 9020B be followed to determine media acceptability prior to analysis.
- “EPA strongly recommends that laboratories evaluate the false-positive and negative rates for method(s) they use for monitoring total coliforms. . . . with the intent that if the method they choose has an unacceptable false-positive or negative rate, another method can be used.” – 40 CFR 141.21 f.3.12
- To avoid reporting any false positive results, follow the coliform verification steps as indicated in the method you are using.
- This standard contains viable microorganisms and should be analyzed **immediately** after being hydrated.
- When performing Multiple-Tube Fermentation, run a series of tubes that would yield a result consistent with the manufacturing range of 20 to 2400 MPN index/100 mL.

#### Instructions:

1. Remove the vials from refrigeration and allow to warm to room temperature.
2. Carefully open the hydrating fluid that has been stored at room temperature.
3. One at a time, open the bacteria sample vials and aseptically transfer the gelatin tablets into the hydrating fluid.
4. Properly dispose of the empty glass vials and desiccant pouches.
5. Reseal the bottles that now contain the bacteria samples.
6. With the bottle tightly closed, shake the sample for a few seconds. Observe the sample to confirm that the gelatin tablet has dissolved. If the tablet has not completely dissolved, shake for a few more seconds.
7. Analyze the inoculated samples using your normal procedure.

#### Safety:

- ERA products may be hazardous and are intended for use by professional laboratory personnel trained in the competent handling of such materials. Responsibility for the safe use of these products rests entirely with the buyer and/or user. Safety Data Sheets (SDS) for all ERA products are available through our website. [www.eraqc.com](http://www.eraqc.com)
- **ERA microbiology standards contain live microorganisms** and should be used only by individuals with bacteriological training.
- Properly disinfect any spills and sterilize used containers by autoclaving before disposal.

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## Microbiology Technical Guide to Getting it Right the First Time

### Introduction:

Before you start the analysis of your quantitative WasteWatR™ Coliforms Quality Control sample (Cat. # 083A) please read through this guide. This document is intended to walk you through all the important steps and ask all the right questions to help ensure that your technique and procedures are correct. This guide is not intended to supersede any specific Quality Assurance Plan or method-specific quality control that you are currently doing. It is only a supplement intended to get you thinking about how you currently perform the method and point out aspects of the process that we at ERA have observed to be common causes of poor method performance. And if you need any help at any step of the way, don't hesitate to call ERA for technical assistance at 1-800-372-0122; our Microbiology Team is here to help!

### General Quality Control:

**Sterility Checks** - Laboratories should perform sterility checks for the procedure in use. For membrane filter procedures check sterility of media, membrane filters, buffered dilution water, pipets, flasks and dishes, and all equipment at the beginning and end of each series of samples, using buffered dilution water as the sample. For multiple-tube procedures, check sterility of media, buffered dilution water, and glassware. Each new batch of the above specific materials should be checked prior to use.

**Positive Controls** - For each new batch of media and method apparatus, check analytical procedure by testing a positive control to demonstrate that the media produces the expected reaction to the organism under test. In order to ensure identity and traceability, reference cultures used for positive controls should come from a recognized national collection, organization, or manufacturer recognized by an accrediting authority. These cultures may be a single-use preparation or cultures maintained by procedures that ensure the continued purity and viability of the organism.

**Negative Controls** - For each new batch of media and method apparatus, check analytical procedure by testing a negative control to demonstrate that the media does not demonstrate the typical positive reaction of the target organism. In order to ensure identity and traceability, reference cultures used for negative controls should come from a recognized national collection, organization, or manufacturer recognized by an accrediting authority. These cultures may be a single-use preparation or cultures maintained by procedures that ensure the continued purity and viability of the organism.

**Method Evaluation** - All methods in use in the laboratory should be evaluated for their ability to produce acceptable results prior to first use. For quantitative microbiology methods this can be accomplished by participation in an approved Proficiency Testing program or the analysis of a Quality Control sample.

**Hydration of Pellet** - The initial hydration of the gelatin tablet supplied with ERA samples should be performed using the instructions provided. Please do not deviate from the instructions as the certified values and acceptance intervals are based on customer data and deviations could result in an incorrect outcome.

Once hydration of the pellet is complete, perform your analysis immediately. Waiting more than 30 minutes to perform your analysis could have an affect on results. Gently shake the sample prior to taking aliquots for analysis.

**Culture Media** - Quality of culture media is critical. Never prepare media from raw ingredients if a source of dehydrated media is available. If preparing dehydrated media, follow instructions closely. Always check pH and make any adjustments that are necessary. Always evaluate media prior to first use. Never use media outside of its expiration date and never use media that has not been stored according to the manufacturer's specifications. Confirm that prepared media and dehydrated media's ingredients and proportions match specifications for method in use.

**Dilution and Rinse Water** - Only hydrate the gelatin tablet using the hydrating fluid supplied by ERA. To perform dilutions and apparatus rinsing, do not use DI or distilled water. ERA recommends Phosphate Buffer or Peptone water as it is not uncommon for DI or distilled water to cause inhibitory effects. Reference SM 9050C 20<sup>th</sup> Edition for the preparation of Buffered water or Peptone water.

**Thermometers** - Use thermometers that are graduated in increments appropriate to the analysis being performed. Check the accuracy of thermometers semiannually against a certified NIST thermometer. Use the correction factors, if any, listed on your traceable thermometer's Certificate of Accuracy.

**Water Bath Incubators** - Verify that your water bath maintains the desired test temperature, such as  $44.5^{\circ}\text{C} \pm 0.2^{\circ}\text{C}$ . Keep an appropriate thermometer immersed in the water bath, monitor and record the temperature twice daily at least 4 hours apart. The stability and uniformity of temperature distribution is important. For best operation, equip your water bath with a gable cover, use a water circulator and place incubator in an area where room temperature is maintained between  $16\text{-}27^{\circ}\text{C}$  ( $60\text{-}80^{\circ}\text{F}$ ).

**Air Incubators** - Verify that your incubator maintains the appropriate test temperature, such as  $35^{\circ}\text{C} \pm 0.5^{\circ}\text{C}$ . Also, verify that cold samples are incubated at the test temperature for the required time. Check and record the temperature twice daily for the shelves in use. The thermometer should be submerged in water or glycerine. Place incubator in an area where room temperature is maintained between  $16\text{-}27^{\circ}\text{C}$  ( $60\text{-}80^{\circ}\text{F}$ ). When you open the door of an air incubator the internal temperature can radically change and once the door is closed the temperature does not immediately return to the desired temperature. This time to return to appropriate incubation temperature may be significant, so opening and closing the incubator should be minimized.

#### Method Specific Issues – Multiple-Tube Fermentation:

**Multiple-Tube Fermentation** - Results reported as "> value" will be scored as that value. It is therefore critical that appropriate dilutions are run. We would recommend that QC samples are run at dilutions to simulate the PT scenario.

**Dilution Scheme** - If performing Multiple-Tube Fermentation, run a series of tubes that would yield a result consistent with the manufacturing range of 20-2400 MPN index/100 mL. For example, a series of 15 tubes at 10 mL, 1 mL, and 0.1 mL aliquots will cover an MPN range of <2 to >1600 MPN/100 mL. If desired to cover a wider range of values alter the dilution series.

**Degassing Broth** - To avoid a false positive result due to dissolved gas in the media broth, pre-warm the room-temperature broth tubes for 30 minutes in a  $35^{\circ}\text{C}$  incubator and remove any gas that develops in the fermentation tubes prior to inoculation. A small gas bubble during sample interpretation may indicate insufficient degassing prior to inoculation.

**Broth Concentration** - For an MPN method, confirm that you are using the appropriate broth strength as described in the specific method in use.

#### References:

Standard Methods for the Examination of Water and Wastewater, 20<sup>th</sup> edition.

2003 NELAC Standard, Quality Systems, Appendix D, June 2003.